In a number of cases the activity of pyruvic decarboxylase in breast muscle was also measured, in the same manner as described by Monfoort². The effect of adding TPP was studied simultaneously. The results are given in Table II. As may be seen, the activity of pyruvic decarboxylase is markedly reduced after feeding pyrithiamine. It can almost completely be restored by adding TPP to the homogenates.

Each Warburg flask contained 200 mg breast muscle, homogenized in 0.1 M phosphate buffer pH 6.2, containing 0.001 M magnesiumchloride and made up to a volume of 2.0 ml. 20 μ Mol sodium pyruvate were added after temp. equilibrium. Temperature: 37° C. Gas phase: nitrogen.

Group	Number of animals	PT	Average surv. time	µMol CO ₂ in two hours		
				— TPP	+ 10 γ TPF	
A	4		19 d.	5.1 ± 0.6	6.1 ± 0.7	
\mathbf{B}	4	4.	19 d.	2.6 ± 0.2	5.5 ± 0.2	

These experiments show that the disappearance of TPP from the tissues and the behaviour of the pyruvic decarboxylase after feeding pyrithiamine run a course, closely parallel to that observed in a thiamine deficiency induced by omitting the vitamin from the diet^{2,10}. As yet there seems to be no occasion for presuming that the deficiencies brought about in these two manners are indeed different from one another.

This work forms part of investigations on the metabolism and physiological function of thiamine by H. G. K. Westenbrink and collaborators.

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- ¹ M. Gruber, Biochim. Biophys. Acta, 10 (1953) 136.
- ² C. H. Monfoort, Biochim. Biophys. Acta, 16 (1955) 219.
- ³ D. W. Woolley, J. Biol. Chem., 191 (1951) 43.
- 4 D. W. WOOLLEY AND R. B. MERRIFIELD, Bull. Soc. Chim. Biol., 36 (1954) 1207.
- ⁵ E. P. Steyn-Parvé, Biochim. Biophys. Acta, 14 (1954) 440.
- ⁶ L. R. CERECEDO, M. SOODAK AND A. J. EUSEBI, J. Biol. Chem., 189 (1951) 293;
 - A. J. EUSEBI AND L. R. CERECEDO, Science, 110 (1949) 162.
- ⁷ B. C. P. Jansen and H. G. K. Westenbrink, Acta Brevia Neerl., 3 (1933) 9.
- 8 H. G. K. WESTENBRINK AND E. P. STEYN-PARVÉ, Int. Rev. Vitamin Res., 21 (1950) 461.
- ⁹ S. EICH AND L. R. CERECEDO, J. Biol. Chem., 207 (1954) 295.
- 10 C. H. Monfoort, Biochim. Biophys. Acta, 8 (1952) 389.

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Intracellular distribution of vitamin K in beef liver

An analysis of the intracellular distribution of vitamin K seemed of interest especially in view of the recent evidence that the vitamin is involved in oxidative phosphorylation (Martius and Nitz-Litzow¹). Data on the distribution of the vitamin in liver are presented in this note.

90 g of beef liver were divided into portions and homogenized in a Waring blendor for 15-30 sec. The blended liver was centrifuged by the method of Schneider and Hoogeboom² except that after separation of the mitochondria the remaining supernatant (containing the submicroscopic particles (P) and the soluble (S) fraction) was not further separated. Janus green staining indicated few mitochondria in the supernatant. The three fractions—nuclei (N), mitochondria (M), and supernatant (P-S)—were lyophilized. Nitrogen was determined by the method of Dumas³. Attempts to identify the vitamin in the lyophilized nuclear fraction by paper chromatography (Green and Dam⁴) of a petroleum ether-ethanol extract and by its reaction (Green and Dam⁵) with aluminium oxide were not successful. The fractions were therefore examined

biologically for their vitamin K activity on vitamin K-deficient chicks (DAM, KRUSE AND Søndergaard⁶).

The beef liver mitochondria and supernatant (Table I) contained nearly the same amount of nitrogen as corresponding rabbit fractions (Le Page and Schneider⁷), but the nitrogen content of the nuclear fraction was about twice that of rabbit. This may represent either a species difference or the presence of a large number of unbroken cells in our nuclear fraction due to the short period of homogenizing (POTTER, RECKNAGEL AND HURLBERT8). It is thus possible that the total analysis of vitamin K found in the nuclear fraction is too high.

TABLE I AMOUNTS OF NITROGEN AND VITAMIN K ACTIVITY EXPRESSED AS MENADIONE IN FRACTIONS OF 100 g FRESH BEEF LIVER

<u>.</u>	N per 100 g of fresh liver	N % of total N	y menadione in fraction of 100 g liver	menadione in % of total menadione	γ menadione per g N of fraction	µM vitamin I per g N of fraction
"N"	1.21	43	32	45	27	0.15
''M''	0.28	10	18	25	64	0.38
"P-S"	1.33	47	21	30	16	0.09
Total	2.82	100	71	100		

The vitamin K values shown in Table I are averages of two determinations. The vitamin K activity of 100 g fresh beef liver was equivalent to that of 71 γ menadione. On a nitrogen basis 24 % of vitamin K was associated with the nuclear (N) fraction, 61 % with the mitochondria (M), and 15% with the supernatant (P-S) fraction. The low concentration of vitamin K in the P-Sfraction is in all likelihood a maximum value, as the Waring blendor is known to disintegrate nuclei and mitochondria and disperse their fragments and contents into the supernatant (POTTER,

RECKNAGEL AND HURLBERT⁸, SCHNEIDER AND HOOGEBOOM⁸).

Thus, in liver vitamin K appears to be associated primarily with mitochondria. Its association with both mitochondria and chloroplasts (DAM, GLAVIND AND NIELSEN¹⁰) (analogues of mitochondria) seems to be more than coincidental.

It is of interest to compare the distribution of vitamin K with that of some respiratory catalysts. Thus, in rat liver homogenate riboflavin (Price, Miller and Miller¹¹) is concentrated in the mitochondria and in the supernatant fractions, and cytochrome c is concentrated in the mitochondrial fraction (Schneider and Hoogeboom²). On a nitrogen basis the molar concentration of vitamin K found in our experiments in beef liver mitochondria is about one-third that of cytochrome c and one-fifth that of riboflavin (PRICE, MILLER AND MILLER 11) found in rat liver mitochondria by other investigators.

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- ¹ C. Martius and D. Nitz-Litzow, Biochim. Biophys. Acta, 13 (1954) 152. ² W. C. Schneider and G. H. Hoogeboom, J. Biol. Chem., 183 (1950) 123.
- ³ A. Bömer, A. Juckenack and J. Tillmans, Handbuch der Lebensmittelchemie, Springer Verlag, Berlin, 1935, Vol. 2, p. 580.
- ⁴ J. P. Green and H. Dam, Acta Chem. Scand., 8 (1954) 1341.
- J. P. GREEN AND H. DAM, Acta Chem. Scand., 8 (1954) 1093.
 H. DAM, I. KRUSE AND E. SØNDERGAARD, Acta Physiol. Scand., 22 (1951) 238.
- ⁷ G. A. Le Page and W. C. Schneider, J. Biol. Chem., 176 (1948) 1021.
 ⁸ V. R. Potter, R. O. Recknagel and R. B. Hurlbert, Federation Proc., 10 (1951) 646.
- ⁹ W. C. Schneider and G. H. Hoogeboom, Cancer Research, 11 (1951) 1.
- ¹⁰ H. Dam, J. Glavind and N. Nielsen, Z. physiol. Chem., 265 (1940) 80.

11 J. M. PRICE, E. C. MILLER AND J. A. MILLER, J. Biol. Chem., 173 (1948) 345.

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